

RECEIVED
CENTRAL FAX CENTER

AUG 07 2006


PATENT

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of: John G. Babish, *et al.*
Application No.: 10/789,814
Filing Date: February 27, 2004
Docket Number: 068911-0075
Title: SYNERGISTIC ANTI-INFLAMMATORY
PHARMACEUTICAL COMPOSITIONS AND METHODS
OF USE
Examiner: Shobha Kantamneni
Art Unit: 1617

CERTIFICATE OF TRANSMISSION

I hereby certify that this correspondence is being deposited with the United States Postal Service as First Class Mail in an envelope addressed to MAIL STOP AMENDMENT, Commissioner for Patents, P.O. Box 1450, Alexandria, VA 22313-1450 on the date indicated below.

Date: 08/07/06
Angelo J. Mignanelli
ERIN M. OLSON

MAIL STOP AMENDMENT
Commissioner for Patents
P.O. Box 1450
Alexandria, VA 22313-1450
Sir:

DECLARATION PURSUANT TO 37 C.F.R. § 1.131

I, John G. Babish declare as follows:

1) I am Dr. John G. Babish, Executive Vice President of Metaproteomics, LLC.
I have held this position since August 2002.

BEST AVAILABLE COPY

U.S.S.N. 09/919,506
John G. Babish, *et al.*
1.132 Declaration of John G. Babish
Page 2 of 7

2) I have Doctorate and Masters degrees, respectively, in Biochemistry and Chemistry from Cornell University, as well as a Bachelor degree in Biochemistry from The Pennsylvania State University. A copy of my Curriculum Vitae is attached as Exhibit A.

3) I am also an inventor named in several domestic and foreign patent applications including U.S. Application Nos. 10/141,085; 10/789,814; 10/789,817; 10/988,393; 10/480,145; 10/484,123; 10/881,404; 10/774,048; 10/464,834; 10/234,002 and 09/952,632 and issued foreign and domestic patents, including U.S. Patent Nos. 6,140,063; 5,506,420; 6,629,835; 6,733,793 and 6,908,630.

4) On the basis of 30 years of training and experience, I am an expert in the art of molecular biology, more specifically, that aspect of molecular biology involving signal transduction. I was a faculty member at the College of Veterinary Medicine, Cornell University for 17 years. As Professor of Pharmacology and Toxicology, my research program involved the elucidation of mechanisms by which xenobiotics affect signaling pathways in normal and transformed cells. Using the tools of molecular biology such as monoclonal antibodies, northern and western blotting and enzyme-linked immunoassays, my research program developed cell-based assays for the identification of small molecules directed at inhibiting selected cellular functions. Findings from these studies were used to identify potential anti-viral and anti-neoplastic pharmacophores from natural products. My research has also identified both positive and negative drug-drug and drug-nutrient interactions.


5) I understand that in the course of the February 7, 2006 Office Action during prosecution in the above-captioned application, Examiner Shobha Kantamneni rejected claims 1 – 7 under 35 U.S.C. § 103(a), as allegedly being unpatentable over Kuhrts (US 2004/0137096, PTO-892) for reasons of obviousness. I respectfully submit that the instant invention was conceived prior to the January 9, 2003 filing date of the cited Kuhrts application and diligently researched until the February 27, 2004.

U.S.S.N. 09/919,506
John G. Babish, *et al.*
1.132 Declaration of John G. Babish
Page 3 of 7

6) The basis to assert that the instant invention was invented prior to the Kuhrt reference cited is supported by copies of laboratory notebook pages dated from June 2002 through December 2003 showing research on the synergistic, anti-inflammatory effects of reduced isoalpha acids (RIAA) and isoalpha acids (IAA). Such support documentation is appended herewith as Exhibit B.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Date: 5-14-06


John G. Babish, Ph.D.
Executive Vice President
Metagenics Research Center - Suite 100
9770 44th Ave. NW
Gig Harbor, WA 98332

15% F:\MIS\AA\The-ly\Babish\103605.DOC

U.S.S.N. 09/919,506

John G. Babish, *et al.*

1.132 Declaration of John G. Babish

Page 4 of 7

RECEIVED
CENTRAL FAX CENTER

AUG 07 2006

Exhibit A**BIOGRAPHICAL SKETCH AND BIBLIOGRAPHY**

John G. Babish

Chairperson, BIONexus, Ltd.

Executive Vice President, Metaproteomics Inc.

Education

Institution and Location of Study	Degree	Date Conferred	Field
The Pennsylvania State University, State College, PA	B.S	1968	Biochemistry
Cornell University, Ithaca, NY	M.S	1974	Chemistry
Cornell University, Ithaca, NY	Ph.D.	1976	Biochemistry

Research and Professional Experience

Aug. 2002 – present Executive Vice President of Research & Development, Metaproteomics, Research Laboratories, Ithaca, NY. Metaproteomics develops clinically proven, patented dietary supplements and pharmaceuticals from natural sources. Duties include the design and evaluation of experiments elucidating mechanism of action and biological activity within complex mixtures.

1998 – present (5% Effort) National Coordinator for the USDA Minor Species Drug Program (NRSP-7). The NRSP-7 program is funded by the USDA to provide funds and expertise necessary for the approval of pharmaceuticals used in the treatment of diseases associated with minor crop species. Duties include the coordination of industrial, academic and regulatory resources necessary for protocol development through final drug approval.

1997 – present Co-founder and Chairperson of BIONexus, Ltd. Ithaca, NY. BIONexus develops and markets nutritional supplements to address health problems associated with AIDS. NutriVir™, the BIONexus supplement for wasting in HIV/AIDS, generated approximately \$600,000 in gross revenues in its first year of sales. NutriVir™ is reimbursed by Medicaid in 14 states.

U.S.S.N. 09/919,506

John G. Babish, *et al.*

1.132 Declaration of John G. Babish

Page 5 of 7

- 1991 – 1996 Founder, Chairperson, President and CEO of Paracelsian, Inc., Ithaca, NY. The Company was launched from the technology transfer program of Cornell University in 1991, and with the public offering in 1992 (Nasdaq:PRLN), became the first public corporation of a Cornell University technology transfer effort. Babish was associated with the attainment of over \$12 million dollars in corporate financing.
- 1984 – 1996 Tenured, Associate and Professor of Pharmacology and Toxicology, Department of Pharmacology, College of Veterinary Medicine, Cornell University. Offered the first course in molecular risk assessment in the USA in 1979; member of the graduate Fields of Pharmacology, Toxicology, Veterinary Medicine, Food Science and Epidemiology; successfully petitioned the State of New York for the approval of the separate Fields of Toxicology and Pharmacology at Cornell University.
- 1978 – 1984 Assistant Professor, Department of Preventive Medicine, NYS College of Veterinary Medicine, Cornell University, Ithaca, NY.
- 1976 – 1978 Postdoctoral Scientist, Food and Drug Research Labs, Waverly, NY.

Invited Presentations (Recent of 38 presentations)

Micronutrient deficiencies in AIDS wasting at Progressive Management of AIDS Wasting: 2000. Hunter College, NYC. March 24, 2000.

Phytochemicals and NF- κ B activation at IBC's Conference on The Health Benefits of Natural Phytochemicals. Montreal Bonaventure Hilton, July 22 – 23, 1997.

Chemically-induced cell cycle stasis in immunotoxicology. 12th Annual NIOSH Conference on Mechanisms of Immunotoxicology – Role of Apoptosis in Immunotoxicology. University of West Virginia, Morgantown, WV. September 10 – 12, 1997.

Publications (Selected of 108 peer-reviewed publications)

Payne M.A., Babish J.G., Bulgin M., Lane M., Wetzlich S., Craigmill A.L. (2002) Serum pharmacokinetics and tissue and milk residues of oxytetracycline in goats following a single intramuscular injection of a long-acting preparation and milk residues following a single subcutaneous injection. *J Vet Pharmacol Ther.* 25(1):25-32.

Calabrese C., Berman S.H., Babish J.G., et al. (2000) A phase I trial of andrographolide in HIV positive patients and normal volunteers. *Phytother Res.* 14(5):333-338.

Ma, X., Stoffregen, D.A., Wheelock, G.D., Rininger, J.A. and Babish, J.G. (1997) Discordant hepatic expression of the cell division control enzyme p34cdc2 kinase, proliferating cell nuclear antigen, p53 tumor suppressor protein, and p21Waf1 cyclin-

U.S.S.N. 09/919,506

John G. Babish, *et al.*

1.132 Declaration of John G. Babish

Page 6 of 7

dependent kinase inhibitory protein after WY14,643 ([4-chloro-6-(2,3-xylidino)-2-pyrimidinylthio]acetic acid) dosing to rats. *Mol. Pharmacol.*, 51, 69-78.

Rininger, J.A., Goldsworthy, T.L. and Babish, J.G. (1997) Time course comparison of cell-cycle protein expression following partial hepatectomy and WY14,643-induced hepatic cell proliferation in F344 rats. *Carcinogenesis*, 18, 935-941.

Rininger, J.A., Stoffregen, D.A. and Babish, J.G. (1997) Murine hepatic p53, RB, and CDK inhibitory protein expression following acute 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) exposure. *Chemosphere*, 34, 1557-1568.

Rininger, J.A., Wheelock, G.D., Ma, X. and Babish, J.G. (1996) Discordant expression of the cyclin-dependent kinases and cyclins in rat liver following acute administration of the hepatocarcinogen [4-chloro-6-(2,3-xylidino)-2-pyrimidinylthio] acetic acid (WY14,643). *Biochem. Pharmacol.*, 52, 1749-1755.

Vancutsem, P.M. and Babish, J.G. (1996) In vitro and in vivo study of the effects of enrofloxacin on hepatic cytochrome P-450. Potential for drug interactions. *Vet. Hum. Toxicol.*, 38, 254-259.

Patents (Selected of 15 US and three foreign patents)

US Patent No. 5,833,994	11/10/1998 Use of the Ah receptor and Ah receptor ligands to treat or prevent cytopathicity of viral infection.
US Patent No. 5,612,188	3/18/1997 Automated, multicompartmental cell culture system.
US Patent No. 5,529,899	6/25/1996 Immunoassay for Ah receptor transformed by dioxin-like compounds.
US Patent No. 5,496,703	3/5/1996 Indirect immunoassay for dioxin-like compounds

U.S.S.N. 09/919,506
John G. Babish, *et al.*
1.132 Declaration of John G. Babish
Page 7 of 7

Exhibit B

LABORATORY NOTEBOOK SHEETS DOCUMENTING RESEARCH ON RIAA, IAA AND OTHER HOPS
DERIVATIVES

Date	Notebook Number	Pages
6/4/02 - 8/24/02	2002-03	13 - 23
8/28/02	2002-04	3 - 4
12/5/02	2002-06	1 - 2
3/5/03	2002-08	43 - 44
4/23/03	2003-01	23
6/5/03	2003-01	45
9/3/03	2003-4	22
12/15/03	2003-5	42

PROJECT POE₂ Assay in RAW cellsNotebook No. 2002-03

Continued From Page

13

Experiment 2002-03-13

The general purpose + procedure can be found on pages 4, 8, and 11 of this notebook

1_C00X-DRAW_0.04.02

Compounds for Metagenesis
POE₂ assay, using 1:10 dilution from the plates
6.04.02 (USING RAW 264.7 cells with LPS stimulation and 15 min with ARA)

Compound	Conc.	Vol.	Conc.	Vol.	Conc.	Vol.	Conc.	Vol.
1. BetaTech - Alpha hop (10 µg/L)	Alpha hop =	50.000	10.000	5.000	1.000	0		
2. BetaTech - Beta acid solution (10 µg/L)	Beta acid solution =	50.000	10.000	5.000	1.000	0		
3. BetaTech Aromashop OE (10 µg/L)	Aromashop OE =	50.000	10.000	5.000	1.000	0		
4. BetaTech Radshop (RAA) (10 µg/L)	Radshop (RAA) =	50.000	10.000	5.000	1.000	0		
5. BetaTech Radshop (RAA) (10 µg/L)	Radshop (RAA) =	50.000	10.000	5.000	1.000	0		
6. BetaTech Tetrahop Gold (10 µg/L)	Tetrahop Gold =	50.000	10.000	5.000	1.000	0		
7. BetaTech HexaShop gold (HRAA) (10 µg/L)	HexaShop gold =	50.000	10.000	5.000	1.000	0		
8. BetaTech Hop oil (10 µg/L)	Hop oil =	50.000	10.000	5.000	1.000	0		
9. Diisopropyl fluorophosphate	DIFP =	50.000	5.000	0.500	0.050	0		
10. Indinavir IC Tablets	Indinavir IC =	50.000	25.000	12.500	4.250	0		
					total =	80		

2_C00X-DRAW_0.04.02

New Hops Compounds for Metagenesis
POE₂ assay, using 1:10 dilution from the plates
6.04.02 (USING RAW 264.7 cells with LPS stimulation and 15 min with ARA)

Compound	Conc.	Vol.	Conc.	Vol.	Conc.	Vol.	Conc.	Vol.
1. BetaTech Radshop (RAA) (10 µg/L)	RAA =	50.000	10.000	5.000	1.000	0		
2. RAAUric acid (1:1)	Total =	50.000	10.000	5.000	1.000	0		
	RAA =	25.000	1.000	2.500	0.500			
	Uric acid =	25.000	1.000	2.500	0.500			
3. Lemon Bioflavonoid (RMD77810)	Bioflavonoid =	50.000	10.000	5.000	1.000	0		
4. Ginger (RMD7782)	Ginger =	50.000	10.000	5.000	1.000	0		
5. RAACurcumin (RMD7782) (1:1)	Total =	50.000	10.000	5.000	1.000	0		
	RAA =	25.000	5.000	2.500	0.500			
	Curcumin =	25.000	5.000	2.500	0.500			
6. Capsaicin Pepper (RMD7782)	Capsaicin =	50.000	10.000	5.000	1.000	0		
7. RAAQuercetin (RMD7781) (1:1)	Total =	50.000	10.000	5.000	1.000	0		
	RAA =	25.000	5.000	2.500	0.500			
	Quercetin =	25.000	5.000	2.500	0.500			
8. RAAAlphit galanga (1:1)	Total =	50.000	10.000	5.000	1.000	0		
	RAA =	25.000	5.000	2.500	0.500			
	Alphit galanga =	25.000	5.000	2.500	0.500			
9. RAACurcumin (RMD7781) (1:1)	Total =	50.000	10.000	5.000	1.000	0		
	RAA =	25.000	5.000	2.500	0.500			
	Curcumin =	25.000	5.000	2.500	0.500			
10. Hennaide	Hennaide =	50.000	10.000	5.000	1.000	0		
					total =	80		

Continued on Page 14

Read and Understood By

Signed

Date

Signed

Date

PROJECT

BEJ In RAW cells

Notebook No 2002-03

Continued From Page

16

Experiment 2002-03 -16

The purpose and procedure for this experiment can be found on pages 4, 8, and 16 of this notebook.

1. CON-2-RAW_8.10.02

Compounds for Metagenesis
PGE2 assay, using 1:10 dilution from the plates
8.10.02 USING RAW 264.7 cells with LPS stimulation and 15 min with ARAI. Dose media 1:50 and 1:100 for PGE2 assay

Compound	Conc	10,000	10,000	5,000	1,000	0
1. BetaTech - Alpha hop (10µg/dL)	Alpha hop =	50,000	10,000	5,000	1,000	0
2. BetaTech - Beta acid solution (10µg/dL)	Beta acid solution =	50,000	10,000	5,000	1,000	0
3. BetaTech Aromahop OE (10µg/dL)	Aromahop OE =	50,000	10,000	5,000	1,000	0
4. BetaTech Isachop (IAA) (10µg/dL)	Isachop (IAA) =	50,000	10,000	5,000	1,000	0
5. BetaTech Radhop (RAA) (10µg/dL)	Radhop (RAA) =	50,000	10,000	5,000	1,000	0
6. BetaTech Tetrahop Gold (10µg/dL)	Tetrahop Gold =	50,000	10,000	5,000	1,000	0
7. BetaTech Hexahop gold (HMAA) (10µg/dL)	Hexahop gold =	50,000	10,000	5,000	1,000	0
8. Ibuprofen	Ibuprofen =	50,000	10,000	5,000	1,000	0
9. Celebrex	Celebrex =	5,000	0,500	0,050	0,005	0
10. Intraenoid IC	Intraenoid IC =	50,000	10,000	5,000	1,000	0
					total =	80

BIO-TEK MICROPLATE READER

06/14/02 AT 01:02 PM

00011864

ASSAY

PLATE

OPERATOR

NOTES

PROGRAM MODE #9

SINGLE WAVELENGTH: 495

TABLE OF ABSORBANCE VALUES

	1	2	3	4	5	6	7	8	9	10	11	12
A	0.887	0.490	0.591	0.541	0.576	0.517	0.529	0.546	0.542	0.697	0.667	0.670
B	1.628	0.533	0.586	0.529	0.571	0.533	0.521	0.526	0.544	0.683	0.609	0.704
C	0.193	0.666	0.519	0.582	0.479	0.515	0.534	0.548	0.584	0.397	0.662	0.598
D	0.163	0.637	0.533	0.514	0.516	0.531	0.527	0.563	0.521	0.509	0.659	0.388
E	0.568	0.578	0.599	0.577	0.562	0.581	0.576	0.616	0.638	0.615	0.649	0.573
F	0.667	0.586	0.514	0.513	0.533	0.534	0.537	0.536	0.526	0.552	0.648	0.554
G	0.156	0.673	0.523	0.532	0.544	0.546	0.588	0.549	0.533	0.596	0.604	0.602
H	0.103	0.653	0.519	0.573	0.519	0.547	0.517	0.548	0.546	0.531	0.638	0.534

BIO-TEK MICROPLATE READER

06/14/02 AT 01:03 PM

00011865

ASSAY

PLATE

OPERATOR

NOTES

PROGRAM MODE #9

SINGLE WAVELENGTH: 495

TABLE OF ABSORBANCE VALUES

	1	2	3	4	5	6	7	8	9	10	11	12
A	0.988	0.641	0.682	0.681	0.659	0.590	0.628	0.621	0.627	0.708	0.694	0.728
B	1.881	0.628	0.553	0.578	0.628	0.682	0.594	0.685	0.624	0.698	0.782	0.732
C	0.168	0.639	0.566	0.535	0.531	0.553	0.586	0.578	0.598	0.638	0.664	0.657
D	0.163	0.627	0.584	0.531	0.572	0.577	0.583	0.606	0.598	0.612	0.678	0.633
E	0.678	0.497	0.633	0.612	0.617	0.648	0.625	0.626	0.598	0.649	0.663	0.661
F	0.593	0.596	0.589	0.585	0.564	0.598	0.577	0.609	0.682	0.628	0.658	0.622
G	0.154	0.673	0.625	0.598	0.588	0.573	0.595	0.584	0.638	0.631	0.783	0.662
H	0.132	0.688	0.646	0.648	0.628	0.631	0.638	0.635	0.782	0.616	0.783	0.628

The same plate was run @ 2 different dilutions. 1:50 and 1:100

Continued on Page

Read and Understood By

Signed

Date

Signed

Date

PROJECT POE Assay + NO AssayNotebook No. 2002-03

Continued From Page

17

Experiment 2002-03-17

- The general purpose and procedure for the POE Assay can be found on 2002-03-04 experiment.
- Purpose for the NO experiment to determine the levels of Nitrite Oxide being produced in the cells.
- Procedure for NO.

Griess Reagent was made.
 1% - Sulfanilamide Sigma 5-9251 lot 101K2010
 0.1% - N-(1-naphthyl) ethylenediamine (Sigma N-5889)
 2.5% - H₃PO₄ (Riedel-de Haen -0410) lot 71150112
 Lot 52540

This reagent is mixed up in Ultra Pure Water
 25 ml = 250 mg Sulfanilamide
 25 mg N-(1-naphthyl) ethylenediamine
 625 mg H₃PO₄

The same plates of cells were used for each experiment.

The cells were taken from 1 X T₇₅ plate of RAW cells that were 80% confluent and plated into 2 96 well plates at 4×10^5 cells per well.

Continued on Page 18

Read and Understood By

[Signature] 6-17-02
 Signed Date

[Signature] 7-19-02
 Signed Date

PROJECT PGC₂ + NO in RAN cellsNotebook No. 2002-03Continued From Page 17

18

2002-03-15/17 cont.

1. CON-RAW_6.17.02

Compounds for Metagenics

PGC₂ assay, using undiluted and 1:1 dilution from the plates6.17.02 USING RAW 264.7 cells with LPS stimulation and 15 min with 5 μ M ARA1

Compounds	1	2	3	4	5
1. BetaTech - Alpha hop (10 μ g/L)	Alpha hop = 50,000	10,000	5,000	1,000	8
2. BetaTech - Beta acid solution (10 μ g/L)	Beta acid solution = 50,000	10,000	5,000	1,000	8
3. BetaTech Aromashop OE (10 μ g/L)	Aromashop OE = 50,000	10,000	5,000	1,000	8
4. BetaTech Isoshop (IAA) (10 μ g/L)	Isoshop (IAA) = 50,000	10,000	5,000	1,000	8
5. BetaTech Radshop (RIAA) (10 μ g/L)	Radshop (RIAA) = 50,000	10,000	5,000	1,000	8
6. BetaTech Tetrahop Gold (10 μ g/L)	Tetrahop Gold = 50,000	10,000	5,000	1,000	8
7. BetaTech Hexashop gold (HIAA) (10 μ g/L)	Hexashop gold = 50,000	10,000	5,000	1,000	8
8. Ibuprofen	Ibuprofen = 50,000	10,000	5,000	1,000	8
9. Celebrex	Celebrex = 5,000	0,500	0,050	0,005	8
10. Inflavonoid IC Tablets	Inflavonoid IC = 50,000	10,000	5,000	1,000	8
				total =	80

The compounds used on these two plates are identical. The amount of Arachidonic Acid used is not.

4 plates will be used for PGE₂ and

Three for NO

2. CON-RAW_6.17.02

Compounds for Metagenics

PGC₂ assay, using undiluted and 1:1 dilution from the plates6.17.02 USING RAW 264.7 cells with LPS stimulation and 15 min with 5 μ M ARA1

Compounds	1	2	3	4	5
1. BetaTech - Alpha hop (10 μ g/L)	Alpha hop = 50,000	10,000	5,000	1,000	8
2. BetaTech - Beta acid solution (10 μ g/L)	Beta acid solution = 50,000	10,000	5,000	1,000	8
3. BetaTech Aromashop OE (10 μ g/L)	Aromashop OE = 50,000	10,000	5,000	1,000	8
4. BetaTech Isoshop (IAA) (10 μ g/L)	Isoshop (IAA) = 50,000	10,000	5,000	1,000	8
5. BetaTech Radshop (RIAA) (10 μ g/L)	Radshop (RIAA) = 50,000	10,000	5,000	1,000	8
6. BetaTech Tetrahop Gold (10 μ g/L)	Tetrahop Gold = 50,000	10,000	5,000	1,000	8
7. BetaTech Hexashop gold (HIAA) (10 μ g/L)	Hexashop gold = 50,000	10,000	5,000	1,000	8
8. Ibuprofen	Ibuprofen = 50,000	10,000	5,000	1,000	8
9. Celebrex	Celebrex = 5,000	0,500	0,050	0,005	8
10. Inflavonoid IC Tablets	Inflavonoid IC = 50,000	10,000	5,000	1,000	8
				total =	80

- 1 - Alpha hop - AN1124
 2 - Beta Acid - AN1125
 3 - Aromashop - AN1126
 4 - Isoshop - AN1127
 5 - Radshop - AN1128
 6 - Tetrahop - AN1128
 7 - Hexashop - AN1129
 8 - Ibuprofen - Sigma I-4683 lot 26 H1368
 9 - Celebrex - AN1055
 10 - Inflavonoid IC Tablets - AN1101

Continued on Page 19

[Signature]
 Signed

6.19.02
6.15.02
 Date

Read and Understood By

[Signature]
 Signed

7-19-02
 Date

PROJECT PGE₂ + NO₂ in RAW cellsNotebook No. 2002-03Continued From Page 19

20

Experiment 2002-03-17 cont.

COX-1 1.2

Bio-Tek Instruments

Assay: Quick Read

Date: 06/30/03

Lot:

Well length: 405

Time: 02:35:43PM

Operator:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

Plate ID:

Assay: 405

Temp:

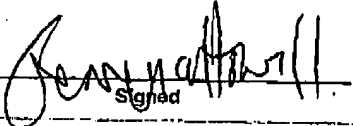
Plate ID:

This is the data from the RAW plate COX-1 1:2 dilution.

This is data from the Undiluted COX-1 plate.

Continued on Page 21

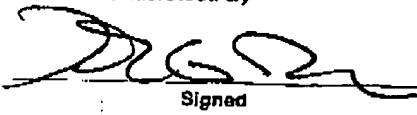
Read and Understood By



Signed

6-20-07

Date



Signed

7-19-02

Date

PROJECT PGE₂ + NO in RAW cells

Notebook No. 780293
 Continued From Page 20

Continued From Page 10

21

Experiment 2002-03-17 cont

A standard curve was done on the NO.

50ml of media Δ of the treated cells
was added to 50ml of the Greiss reagent
This was allowed to sit for 10 minutes
then read @ 605 nm.

No St. Curve

This is the NO
Standard curve

b6-Tek Instruments

ANSWER: Quick Read

Date: 08/20/02

Let

Month: 6/10

7/4/2011 10:28:22 AM

Operator:

1115-1116

915- 10,
104444244

CONCLUSIONS

[illegible]

Continued on Page 22

Read and Understood By

Signed _____

Signed

620-02
Date

Date

DGC

Signed

7-19-02

Date _____

PROJECT PE₂ - No Assay in RAW cellsNotebook No. 2002-03Continued From Page 21

22

Experiments 2002-03-17

Bio-Tek Instruments

Assay: Quick Read

Date: 08/20/02

Lot:

Well length: 400

Time: 02:33:40 PM

Operator:

Temp: _____

Plate ID:

CONCENTRATIONS

1	2	3	4	5	6	7	8	9	10	11	12	13
ChL	0.125	0.116	0.107	0.098	0.089	0.080	0.071	0.062	0.053	0.044	0.035	0.026
ChL	0.025	0.023	0.021	0.019	0.017	0.015	0.013	0.011	0.009	0.007	0.005	0.003

1	2	3	4	5	6	7	8	9	10	11	12	13
ChL	0.125	0.116	0.107	0.098	0.089	0.080	0.071	0.062	0.053	0.044	0.035	0.026
ChL	0.025	0.023	0.021	0.019	0.017	0.015	0.013	0.011	0.009	0.007	0.005	0.003

1	2	3	4	5	6	7	8	9	10	11	12	13
ChL	0.125	0.116	0.107	0.098	0.089	0.080	0.071	0.062	0.053	0.044	0.035	0.026
ChL	0.025	0.023	0.021	0.019	0.017	0.015	0.013	0.011	0.009	0.007	0.005	0.003

1	2	3	4	5	6	7	8	9	10	11	12	13
ChL	0.125	0.116	0.107	0.098	0.089	0.080	0.071	0.062	0.053	0.044	0.035	0.026
ChL	0.025	0.023	0.021	0.019	0.017	0.015	0.013	0.011	0.009	0.007	0.005	0.003

Bio-Tek Instruments

Assay: Quick Read

Date: 08/20/02

Lot:

Well length: 400

Time: 02:38:07 PM

Operator:

Temp: _____

Plate ID:

CONCENTRATIONS

1	2	3	4	5	6	7	8	9	10	11	12	13
ChL	0.125	0.116	0.107	0.098	0.089	0.080	0.071	0.062	0.053	0.044	0.035	0.026
ChL	0.025	0.023	0.021	0.019	0.017	0.015	0.013	0.011	0.009	0.007	0.005	0.003

1	2	3	4	5	6	7	8	9	10	11	12	13
ChL	0.125	0.116	0.107	0.098	0.089	0.080	0.071	0.062	0.053	0.044	0.035	0.026
ChL	0.025	0.023	0.021	0.019	0.017	0.015	0.013	0.011	0.009	0.007	0.005	0.003

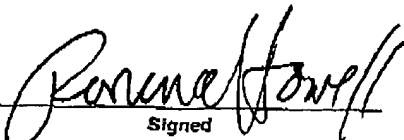
1	2	3	4	5	6	7	8	9	10	11	12	13
ChL	0.125	0.116	0.107	0.098	0.089	0.080	0.071	0.062	0.053	0.044	0.035	0.026
ChL	0.025	0.023	0.021	0.019	0.017	0.015	0.013	0.011	0.009	0.007	0.005	0.003

1	2	3	4	5	6	7	8	9	10	11	12	13
ChL	0.125	0.116	0.107	0.098	0.089	0.080	0.071	0.062	0.053	0.044	0.035	0.026
ChL	0.025	0.023	0.021	0.019	0.017	0.015	0.013	0.011	0.009	0.007	0.005	0.003

COX-2 RAW data
NOCOX-1 RAW data
NO

Continued on Page

Read and Understood By



Signed

6-20-02
Date



Signed

7-19-02
Date

PROJECT PGC₂ in RAW cells + NO AssayNotebook No. 2002-03

Continued From Page

23

Experiment 2002-03-23

The purpose and procedure can be found on page 11 of this notebook.

1_COX-2RAW_8.24.02

Compounds for Metagenics

PGC₂ assay, using 1:1 and 1:10 dilution from the plates6.24.02 USING RAW 264.7 cells with LPS stimulation and 15 min with 5 μ M ARA

Compound	Concentration	50.000	5.000	0.500	0.050	8
1. Alcohol extract of spent hops	EXHSH =	50.000	5.000	0.500	0.050	8
2. Urolic acid 90% - Salinas	Urolic acid =	50.000	5.000	0.500	0.050	8
3. Catechol acid (Rosemary)	Salutic acid =	50.000	5.000	0.500	0.050	8
4. Catechol acid 80% - Salinas	Catechol acid 80% =	50.000	5.000	0.500	0.050	8
5. BetaTech Radhop (RAA) (10 μ g/L)	Radhop (RAA) =	50.000	5.000	0.500	0.050	8
6. Curcumin granular (9855)	Curcumin =	50.000	5.000	0.500	0.050	8
7. RAA:Urolic acid (90%) (1:1)	Total =	50.000	5.000	0.500	0.050	8
	RAA =	25.000	2.500	0.250	0.025	8
	Urolic acid 90% =	25.000	2.500	0.250	0.025	8
8. Ibuprofen - Sigma	Ibuprofen =	50.000	5.000	0.500	0.050	8
9. Celebrex	Celebrex =	5.000	0.500	0.050	0.005	8
10. Aspirin - Sigma	Aspirin =	50.000	10.000	5.000	1.000	8
	total =				80	

2_COX-1RAW_8.24.02

Compounds for Metagenics

PGC₂ assay, using undiluted and 1:1 dilution from the plates6.24.02 USING RAW 264.7 cells no LPS stimulation and 60 min with 100 μ M ARA

Compound	Concentration	50.000	5.000	0.500	0.050	8
1. Alcohol extract of spent hops		50.000	5.000	0.500	0.050	8
2. Urolic acid 90% - Salinas		50.000	5.000	0.500	0.050	8
3. Catechol acid (Rosemary)		50.000	5.000	0.500	0.050	8
4. Catechol acid 80% - Salinas		50.000	5.000	0.500	0.050	8
5. BetaTech Radhop (RAA) (10 μ g/L)	Radhop (RAA) =	50.000	5.000	0.500	0.050	8
6. Curcumin granular (9855)		50.000	5.000	0.500	0.050	8
7. RAA:Urolic acid (90%) (1:1)	Total =	50.000	5.000	0.500	0.050	8
	RAA =	25.000	2.500	0.250	0.025	8
	Urolic acid 90% =	25.000	2.500	0.250	0.025	8
8. Ibuprofen - Sigma	Ibuprofen =	50.000	5.000	0.500	0.050	8
9. Celebrex	Celebrex =	5.000	0.500	0.050	0.005	8
10. Aspirin - Sigma	Aspirin =	50.000	10.000	5.000	1.000	8
	total =				80	

Continued on Page 24

Read and Understood By



Signed

6.24.02

Date



Signed

7-19-02

Date

PROJECT PGE₂ Expression in AGS+ A-549 cellsNotebook No. 2002-04Continued From Page 22002-0304-01 cont.

C2_COX2_A549_8.22.02

A549 Cells - wash, cells treated with test material, stimulated with IL-1/LPS/IL-2 for 24 hrs and assayed for PGE₂.
For PGE₂ assay see media modified and diluted 1:20.

	IAA =	25	5.0	0.5	0.05	0
1. DataTech (IAA)	DataTech (IAA) =	25	5.0	0.5	0.05	0
2. DataTech (RIA)	Total =	25	5.0	0.5	0.05	0
3. IAA:RIA (1:1)	IAA =	12.5	2.5	0.25	0.025	0
	RIA =	12.5	2.5	0.25	0.025	0
4. IAA:RIA (2:1)	Total =	25	5	0.5	0.05	0
	IAA =	16.7	3.3	0.33	0.033	0
	RIA =	8.3	1.7	0.167	0.017	0
5. IAA:RIA (5:1)	Total =	25	5	0.5	0.05	0
	IAA =	20.0	4.0	0.4	0.04	0
	RIA =	5.0	1.0	0.1	0.01	0
6. IAA:RIA (10:1)	Total =	25	5	0.5	0.05	0
	IAA =	22.7	4.55	0.455	0.0455	0
	RIA =	2.3	0.45	0.045	0.0045	0
7. IAA:RIA (50:1)	Total =	25	5	0.5	0.05	0
	IAA =	24.0	4.80	0.480	0.0480	0
	RIA =	1.0	0.20	0.020	0.0020	0
8. IAA:RIA (100:1)	Total =	25	5	0.5	0.05	0
	IAA =	24.75	4.95	0.495	0.0495	0
	RIA =	0.25	0.05	0.005	0.0005	0
9. IAA:Trypanthine (1:1)	Total =	25	5	0.5	0.05	0
	IAA =	12.5	2.5	0.25	0.025	0
	Trypanthine =	12.5	2.5	0.25	0.025	0
	Total =	72				

Note 1000, 500, 10.0 and 1.0 concentrations of the standard curve are listed in Column 2

Same compounds as
on page 20 of this
notebookThis plate was treated the same as the plate on
page 2 of this notebook

C3_COX2_A549_8.22.02D6

A549 Cells - cells are stimulated with IL-1/LPS/IL-2 for 24 hrs, washed, test materials added for 60 minutes then A23187 (50 µM) added as
assay for PGE₂ 30 minutes later.
For PGE₂ assay see media modified and diluted 1:20
8.22.02

1. Diisopropyl fluorophosphate	COX-2	25	5.0	0.5	0.05	0
2. Vinore	COX-2	25	5.0	0.5	0.05	0
3. Celecox	COX-2	25	5.0	0.5	0.05	0
4. Nimesulide	COX-2	25	5.0	0.5	0.05	0
5. Ibuprofen	COX-2/COX-1	25	5.0	0.5	0.05	0
6. Indomethacin	COX-1	25	5.0	0.5	0.05	0
7. Aspirin	COX-1	25	5.0	0.5	0.05	0
8. Salicylic acid	-	25	5.0	0.5	0.05	0
9. Naproxen	-	25	5.0	0.5	0.05	0
10. Acetaminophen	-	25	5.0	0.5	0.05	0
	Total =	80				

1- Aldrich D12/600 lot 0645625
2- AN1066
3- AN1055
4- N-1016 lot 117H1019
5- Sigma I-4863 lot 26H1268
6- Sigma I-7578 lot 60K0745
7- A-5376 lot 119H0175
8- AN1071
9- N-8280 lot 11K1767
10- A-2085 lot 20K068
A-23187 - Sigma
C-7522 lot 81K4002

This plate was stimulated with IL-1/LPS and H₂O₂ for 24 hrs
The plate will then be washed and test materials
added for 1 hr then stimulated with A23187
for 3 minutes

Continued on Page 4

Read and Understood By

Jerome M. Will
Signed

Date

8-28-02

Signed

Jerome M. Will
Signed

Date

9-1-02

PROJECT POE₂ Expression in A-549 CellsNotebook No. 2002-04Continued From Page 3

AGS Cells Format A - wash, add test material and assay for POE₂ the next day
For POE₂ assay run media undiluted
8.24.02

Experiment 2002-04-01

1. Oleic acid (50% Oleic acid)	Oleic acid =	25	5.0	0.5	0.05	0
2. BetaTech (RIAA)	BetaTech (RIAA) =	25	3.0	0.3	0.05	0
3. TrypanBlue - Waco Chemicals	TrypanBlue =	10	1	0.1	0.01	0
4. RIAA:Oleic acid - (1:1)	Total =	25	5	0.5	0.05	0
	RIAA =	22.7	4.5	0.455	0.045	0
	Oleic acid =	2.3	0.5	0.045	0.005	0
5. RIAA:Oleic acid - (5:1)	Total =	25	5	0.5	0.05	0
	RIAA =	20.8	4.2	0.437	0.042	0
	Oleic acid =	4.2	0.8	0.063	0.008	0
6. RIAA:Oleic acid - (1:5)	Total =	25	5	0.5	0.05	0
	RIAA =					
	Oleic acid =					
7. RIAA:Oleic acid - (1:10)	Total =	25	5	0.5	0.05	0
	RIAA =					
	Oleic acid =					
8. #1115 Metoprolol	#1115 =	25	5	0.5	0.05	0
9. RIAA:TrypanBlue - (1:1)	Total =	25	5	0.5	0.05	0
	RIAA =					
	TrypanBlue =					
		Total =	72			

Run 1000, 500, 15.0 and 7.5 concentrations of the standard curve as series in Column 2

AGS Cells Format A - wash, add test material and assay for POE₂ the next day
For POE₂ assay run media undiluted
8.24.02

1. BetaTech (RIAA)	RIAA =	25	5.0	0.5	0.05	0
2. BetaTech (RIAA)	BetaTech (RIAA) =	25	5.0	0.5	0.05	0
3. IAA:RIAA - (1:1)	Total =	25	5.0	0.5	0.05	0
	IAA =	12.5	2.5	0.25	0.025	0
	RIAA =	12.5	2.5	0.25	0.025	0
4. IAA:RIAA - (2:1)	Total =	25	5	0.5	0.05	0
	IAA =	16.7	3.3	0.333	0.033	0
	RIAA =	8.3	1.7	0.167	0.017	0
5. IAA:RIAA - (5:1)	Total =	25	5	0.5	0.05	0
	IAA =	20.8	4.2	0.42	0.042	0
	RIAA =	4.2	0.8	0.08	0.008	0
6. IAA:RIAA - (10:1)	Total =	25	5	0.5	0.05	0
	IAA =	22.7	4.55	0.455	0.0455	0
	RIAA =	2.3	0.45	0.045	0.0045	0
7. IAA:RIAA - (20:1)	Total =	25	5	0.5	0.05	0
	IAA =	24.5	4.9	0.49	0.049	0
	RIAA =	0.5	0.1	0.01	0.001	0
8. IAA:RIAA - (100:1)	Total =	25	5	0.5	0.05	0
	IAA =	24.75	4.95	0.495	0.0495	0
	RIAA =	0.25	0.05	0.005	0.0005	0
9. IAA:TrypanBlue - (1:1)	Total =	25	5	0.5	0.05	0
	IAA =	12.5	2.5	0.25	0.025	0
	TrypanBlue =	12.5	2.5	0.25	0.025	0
		Total =	72			

Run 1000, 500, 15.0 and 7.5 concentrations of the standard curve as series in Column 2

AGS Cells - cells grown to confluence, wash, test materials added for 60 minutes then A23187 (50 µM) added and
assay for POE₂ 30 minutes later.
For POE₂ assay run media undiluted and 120
8.24.02

1. Diisopropyl fluorophosphate	COX-2	25	5.0	0.5	0.05	0
2. Vioxx	COX-2	25	5.0	0.5	0.05	0
3. Celecoxib	COX-2	25	5.0	0.5	0.05	0
4. Nimodipine	COX-2	25	5.0	0.5	0.05	0
5. Ibuprofen	COX-2/COX-1	25	5.0	0.5	0.05	0
6. Indomethacin	COX-1	25	5.0	0.5	0.05	0
7. Aspirin	COX-1	25	5.0	0.5	0.05	0
8. Salicylic acid	-	25	5.0	0.5	0.05	0
9. Naproxen	-	25	5.0	0.5	0.05	0
10. Acetaminophen	-	25	5.0	0.5	0.05	0
		Total =	80			

Plate 3 was done with
the test materials
added and then 60
minutes later they
were stimulated w/
A23187 for 30 minutes
before samples were taken
for POE₂ plates

Continued on Page 5

Read and Understood By

James M. Will
Signed

Date

8-28-02

Dr. G. R.
Signed

Date

9-1-02

PROJECT Cell Lyse + Keko PUG 37Notebook No 2002-06
Continued From Page 1

2

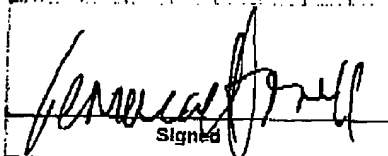
Experiment + 2002-06-01

COX-2 - 56KD

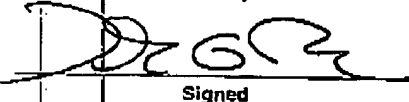
COX-1 66KD

iNOS 130KD

Continued on Page


Signed12-5-02
Date

Read and Understood By


Signed12-23-02
Date

PROJECT

PGE Assay in AGS cells

Notebook No. 2002-06

Continued From Page

43

Experiment 2002-06-43

Purpose + Procedure can be found on pages 2002-06-11

C1_COXAGS_03.03.03

AGS Cells - cells grown to confluence, wash, test materials added for 60 minutes then A23187 (50 μ M) added and assay for PGE2 30 minutes later.
For PGE2 assay run media diluted 1:20 only
3.03.03

Compound	Comments	d1 (μ g/mL)	d2 (μ g/mL)	d3 (μ g/mL)	d4 (μ g/mL)	No Wells
1. Uridine diphosphate		50	5.0	0.5	0.05	8
2. METABOL		50	5.0	0.5	0.05	8
3. Rosemary extract (07720)		50	5.0	0.5	0.05	8
4. Quercetin acid (80% Gabasa)		50	5.0	0.5	0.05	8
5. RIAA (Hess)		50	5.0	0.5	0.05	8
6. Rutin (08284)		50	5.0	0.5	0.05	8
7. Curcumin (07387)		50	5.0	0.5	0.05	8
8. Ginger root (08306)		50	5.0	0.5	0.05	8
9. AHB		50	5.0	0.5	0.05	8
10. Aspirin (Stam)		50	5.0	0.5	0.05	8
Total =						80

C2_COXAGS_03.03.03

AGS Cells - cells grown to confluence, wash, test materials added for 60 minutes then A23187 (50 μ M) added and assay for PGE2 30 minutes later.
For PGE2 assay run media diluted 1:20 only
3.03.03

Compound	Comments	d1 (μ g/mL)	d2 (μ g/mL)	d3 (μ g/mL)	d4 (μ g/mL)	No Wells
1. Isobut AN1127		50	5.0	0.5	0.05	8
2. B1115		50	5.0	0.5	0.05	8
3. Tolshop AN1120		50	5.0	0.5	0.05	8
4. Harschop AN1130		50	5.0	0.5	0.05	8
5. Alphashop AN1124		50	5.0	0.5	0.05	8
6. BetaShop AN1126		50	5.0	0.5	0.05	8
7. Iso-Rich AN1060		50	5.0	0.5	0.05	8
8. Tannin Extract #411 AN1173		50	5.0	0.5	0.05	8
9. A70 LIPOTECH		50	5.0	0.5	0.05	8
10. Aromashop AN1120		50	5.0	0.5	0.05	8
Total =						80

Continued on Page 44

Read and Understood By

Signed

Date

Signed

Date

PROJECT *PEG Army in ASA cells*Notebook No. *2002-05*Continued From Page *43*

44

*Experiment 2002-6-03 con't**RAW Data*

Bio-Tek Instruments

Assay: Quick Read

Date: 08/07/05

Lot: *165 170 plate 1*

Length: 405

Time: 12:47:37PM

Operator:

Plate ID:

Comments

	1	2	3	4	5	6	7	8	9	10	11	12
CHL												
Calcd	0.047	0.138	0.235	0.271	0.379	0.334	0.358	0.347	0.429	0.143	0.201	0.043
Wall	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001
SLT												
CHL												
Calcd	0.037	0.115	0.247	0.257	0.351	0.348	0.371	0.367	0.473	0.173	0.237	0.042
Wall	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001
SLT												
CHL												
Calcd	0.136	0.171	0.145	0.341	0.377	0.352	0.438	0.379	0.429	0.429	0.151	0.104
Wall	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001
SLT												
CHL												
Calcd	0.139	0.182	0.424	0.408	0.371	0.443	0.433	0.431	0.431	0.471	0.043	0.177
Wall	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001
SLT												
CHL												
Calcd	0.137	0.137	0.423	0.423	0.436	0.427	0.423	0.380	0.378	0.432	0.379	0.488
Wall	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001
SLT												
CHL												
Calcd	0.113	0.144	0.413	0.413	0.471	0.477	0.481	0.481	0.447	0.479	0.479	0.487
Wall	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001
SLT												
CHL												
Calcd	0.118	0.171	0.444	0.444	0.449	0.444	0.444	0.444	0.444	0.444	0.444	0.444
Wall	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001
SLT												
CHL												
Calcd	0.169	0.172	0.444	0.479	0.471	0.433	0.433	0.443	0.443	0.457	0.457	0.457
Wall	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001
SLT												

Bio-Tek Instruments

Assay: Quick Read

Date: 08/07/05

Lot: *165 170 plate 1*

Length: 405

Time: 12:52:28PM

Operator:

Plate ID:

Comments

	1	2	3	4	5	6	7	8	9	10	11	12
CHL												
Calcd	0.143	0.114	0.140	0.333	0.321	0.309	0.497	0.372	0.316	0.252	0.252	0.252
Wall	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001
SLT												
CHL												
Calcd	0.712	0.827	0.842	0.378	0.320	0.310	0.363	0.360	0.336	0.321	0.327	0.374
Wall	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001
SLT												
CHL												
Calcd	0.147	0.147	0.379	0.197	0.341	0.343	0.367	0.379	0.361	0.374	0.370	0.367
Wall	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001
SLT												
CHL												
Calcd	0.147	0.147	0.342	0.339	0.337	0.328	0.340	0.370	0.364	0.337	0.343	0.339
Wall	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001
SLT												
CHL												
Calcd	0.154	0.147	0.347	0.340	0.341	0.370	0.371	0.370	0.360	0.379	0.345	0.345
Wall	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001
SLT												
CHL												
Calcd	0.146	0.147	0.333	0.328	0.340	0.372	0.373	0.363	0.360	0.369	0.370	0.368
Wall	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001
SLT												
CHL												
Calcd	0.130	0.134	0.348	0.347	0.348	0.347	0.343	0.337	0.337	0.337	0.337	0.337
Wall	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001
SLT												
CHL												
Calcd	0.147	0.147	0.344	0.344	0.344	0.344	0.344	0.344	0.344	0.344	0.344	0.344
Wall	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.001
SLT												

Continued on Page

Read and Understood By

[Signature]

Signed

3-2-03

Date

[Signature]

Signed

3-9-03

Date

PROJECT AGE₂ Assay in AGS cellsNotebook No. 2003-01

Continued From Page

23

Experiment 2003-01-23

Purpose + Procedure: The purpose is to look at the expression of AGE₂ in AGS cells when exposed to multiple natural compounds.

The procedure can be found on 2003-01-23

4.23.03 - The cells were seeded at 7x10⁵ cells per well in 2-96 well plates.

The following treatments will be used on the AGS cells. AGS cells - ATCC # HTB-79 lot 1641648

AGS Cells - cells grown to confluence, wash, feed medium added for 30 minutes then AZ157 (50 µM) added and assay for AGE₂ 30 minutes later.
For AGE₂ assay run media diluted 1:20 only
5.23.03

Compound	Comments	d1 [µg/mL]	d2 [µg/mL]	d3 [µg/mL]	d4 [µg/mL]	No Wells
1. AGE ₂ Assay (1:1)	Compound = 50 RNA = 25 Aspirin = 25	5.0 2.5 2.5	0.5 0.25 0.25	0.05 0.025 0.025		
2. RNA Assay (1:1)	Compound = 50 RNA = 45.5 Aspirin = 4.5	5.0 4.55 0.5	0.5 0.425 0.075	0.05 0.025 0.025		
3. RNA Assay (1:10)	Compound = 50 RNA = 45.5 Aspirin = 4.5	5.0 4.55 0.5	0.5 0.425 0.075	0.05 0.025 0.025		
4. RNA Assay (1:100)	Compound = 50 RNA = 4.55 Aspirin = 0.455	5.0 4.55 0.455	0.5 0.0455 0.0455	0.05 0.00455 0.00455		
5. RNA Assay (1:1000)	Compound = 50 RNA = 0.455 Aspirin = 0.0455	5.0 0.455 0.0455	0.5 0.00455 0.00455	0.05 0.000455 0.000455		
6. RNA Assay (1:10000)	Compound = 50 RNA = 0.0455 Aspirin = 0.00455	5.0 0.0455 0.00455	0.5 0.000455 0.000455	0.05 0.0000455 0.0000455		
7. RNA Assay (1:100000)	Compound = 50 RNA = 0.00455 Aspirin = 0.000455	5.0 0.00455 0.000455	0.5 0.0000455 0.0000455	0.05 0.00000455 0.00000455		
8. RNA Assay (1:1000000)	Compound = 50 RNA = 0.000455 Aspirin = 0.0000455	5.0 0.000455 0.0000455	0.5 0.00000455 0.00000455	0.05 0.000000455 0.000000455		
9. RNA Assay (1:10000000)	Compound = 50 RNA = 0.0000455 Aspirin = 0.00000455	5.0 0.0000455 0.00000455	0.5 0.000000455 0.000000455	0.05 0.0000000455 0.0000000455		
10. RNA Assay (1:100000000)	Compound = 50 RNA = 0.00000455 Aspirin = 0.000000455	5.0 0.00000455 0.000000455	0.5 0.0000000455 0.0000000455	0.05 0.00000000455 0.00000000455		

C1_COXAGS_04.21.03

AGS Cells - cells grown to confluence, wash, feed medium added for 30 minutes then AZ157 (50 µM) added and assay for AGE₂ 30 minutes later.
For AGE₂ assay run media diluted 1:20 only
5.23.03

Compound	Comments	d1 [µg/mL]	d2 [µg/mL]	d3 [µg/mL]	d4 [µg/mL]	No Wells
1. Isohop AN1127		50	5.0	0.5	0.05	8
2. 61115		50	5.0	0.5	0.05	8
3. Tetrahop AN1120		50	5.0	0.5	0.05	8
4. Hexahop AN1130		50	5.0	0.5	0.05	8
5. Alphahop AN1124		50	5.0	0.5	0.05	8
6. BetaHop AN1125		50	5.0	0.5	0.05	8
7. Iso-Rich AN1090		50	5.0	0.5	0.05	8
8. Tannin Extract 84411 AN1173		50	5.0	0.5	0.05	8
9. AYU LIPOTECH		50	5.0	0.5	0.05	8
10. Avianhop AN1125		50	5.0	0.5	0.05	8
Total =						80

Continued on Page 22

Read and Understood By

Jenniferevill
Signed

4.23.03
Date

[Signature]
Signed

4-26-03
Date

PROJECT Antioxidant Study in HAEC cellsNotebook No. 2003-01

45

Continued From Page

Experiment 2003-01-95Antioxidant Testing in HAEC
Antioxidant assay using HAEC and 1000 μ M H₂O₂+DMSO
Testing dates:

No.	Test Material	6/3/03	6/4/03	6/5/03	No. Wells
		d1 [μ g/mL]	d2 [μ g/mL]	d3 [μ g/mL]	
1	Hopsteiner CO2 Hop Extract (AN1082)	100	10.0	1.00	8
2	Hopsteiner Beta Aroma Extract Light Stable (AN1083)	100	10.0	1.00	8
3	5% Beta Hydrohop (AN1084)	100	10.0	1.00	8
4	5% Alpha Hydrohop (AN1085)	100	10.0	1.00	8
5	YC-Hop Aroma (AN1086)	100	10.0	1.00	8
6	YC-AlphaRich (AN1087)	100	10.0	1.00	8
7	YC-TETRA (AN1088)	100	10.0	1.00	8
8	YC-Kettle RHO (AN1089)	100	10.0	1.00	8
9	ISO-Rich (AN1090)	100	10.0	1.00	8
10	YC-Purified Alpha	100	10.0	1.00	8
total =					80

Purpose + Procedure
203-01-30.

Plate 1

08.02.03/2_HAEC_ROI

10:100
10ul into 90ulAntioxidant Testing in HAEC
Antioxidant assay using HAEC and 1000 μ M H₂O₂+DMSO
Testing dates:

No.	Test Material	6/3/03	6/4/03	6/5/03	No. Wells
		d1 [μ g/mL]	d2 [μ g/mL]	d3 [μ g/mL]	
1	BetaTech 1% AlphaHop (AN1124)	100	10.0	1.00	8
2	BetaTech 1% BetaStab 10A (AN1125)	100	10.0	1.00	8
3	BetaTech 1% AromaHop (AN1126)	100	10.0	1.00	8
4	BetaTech 1% Isohop (AN1127)	100	10.0	1.00	8
5	BetaTech 1% RedHop (AN1128)	100	10.0	1.00	8
6	BetaTech 1% TetraHop Gold (AN1129)	100	10.0	1.00	8
7	BetaTech 1% HexaHop Gold (AN1130)	100	10.0	1.00	8
8	BetaTech 1% Hope Oil (AN1131)	100	10.0	1.00	8
9	Mg Rho 50 (RIAA) (AN1176)	100	10.0	1.00	8
10	RIAA Hop #1199 (AN1177)	100	10.0	1.00	8
total =					80

Plate 2

Continued on Page 46

Signed [Signature]6-5-03
Date

Read and Understood By

Signed [Signature]6-15-03
Date

RAW

Notebook No. 2003-04

22

PROJECT 8/5/03 lysates on Cox-1, Cox-2, iNOS, IK β Blots

Continued From Page

2003-04-22

Purpose: To look at Cox-1, Cox-2, iNOS, IK β in RAW cell lysates from 2003-03-41

Procedure:

4 Gels will be run using the Savant mini-electroblot system. Gels were removed from 4°C and rinsed in ultra-pure water. They were snapped into the savant system and 1x SDS PAGE buffer was added. See 2003-04-08 for 1x SDS PAGE buffer.

4 sets of D. Siml Eppendorf self-locking tubes were set up. Equal volume of loading buffer (Sigma S-3401 lot 21K 5279) and protein sample (volume is in experiment 2003-03-41) were added to appropriate tubes. The tubes were shut and heated @ 100°C for 4 minutes to degrade proteins using a VWR standard HeatBlock. The samples were then spun down briefly using an E.C. micromax ultra-centrifuge.

Then standard was added at 7ul to lane 1 of each gel (BioRad 161-0263 Precision plus Protein standard - unstained)

Lane	2	- control	lot # 96182
"	3	- LPS 10ug/ml	
"	4	- RIAA 10ug/ml	
"	5	- RIAA/LPS 10ug/10ug/ml	
"	6	- Curcumin 10ug/ml	
"	7	- Curcumin/RIA 5ug/ml/5ug/ml	
"	8	- Curcumin/RIA 5ug/ml/5ug/ml	
"	9	- Curcumin/RIA 1LPS 5ug/5ug/10ug/ml	
"	10	- Caffeine 25ug/ml (used from experiment 2003-03-41)	
"	11	- Caffeine/LPS 25ug/10ug/ml	
"	12	- Caffeine/RIA/LPS 5ug/5ug/10ug/ml	
"	13	- note order change!	

Continued on Page 23

Read and Understood By

Signed *James M. Will*

9-3-03

James M. Will

9-5-03

PROJECT DE, Agency in RA + HASCNotebook No. 2003-05
Continued From Page _____

42

2003-05-42Procedure on Page 2003-05-03

C1_COXAGS_12.15.03

AGS Cells - cells grown to confluence, wash, test materials added for 60 minutes then A23187 [50 μ M] added and assay for PGE2 30 minutes later.
For PGE2 assay run media diluted 1:20 only
12/18/03

Compound	Comments	d1 [μ g/mL]	d2 [μ g/mL]	d3 [μ g/mL]	d4 [μ g/mL]	No Wells
1. Salixalin		50	5	0.5	0.05	8
2. New Naprox		50	5	0.5	0.05	8
3. Old Naprox		50	5	0.5	0.05	8
4. Pain Out		50	5	0.5	0.05	8
5. Joint Ease		50	5	0.5	0.05	8
6. Pharma Plam		50	5	0.5	0.05	8
7. Aspirin		50	5	0.5	0.05	8
8. APHB (Cayman)		50	5	0.5	0.05	8
Total =						64

C2_COXAGS_12.16.03

AGS Cells - cells grown to confluence, wash, test materials added for 60 minutes then A23187 [50 μ M] added and assay for PGE2 30 minutes later.
For PGE2 assay run media diluted 1:20 only
12/18/03

Compound	Comments	d1 [μ g/mL]	d2 [μ g/mL]	d3 [μ g/mL]	d4 [μ g/mL]	No Wells
1. Devia Care 600 PLUS		50	5	0.5	0.05	8
2. LYMPHODRAN ORTHOPLEX		50	5	0.5	0.05	8
3. ARLX ORTHOPLEX		50	5.0	0.5	0.05	8
4. ARTHRO-COMPLEX		50	5.0	0.5	0.05	8
5. ARENCAPI®		50	5.0	0.5	0.05	8
6. Traumagel®		50	5.0	0.5	0.05	8
7. Indomethacin		50	5.0	0.5	0.05	8
8. Celebrex®		50	5.0	0.5	0.05	8
Total =						64

Continued on Page 43

Read and Understood By

[Signature]
Signed

12-15-03
Date

[Signature]
Signed

12-20-03
Date

**This Page is Inserted by IFW Indexing and Scanning
Operations and is not part of the Official Record**

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images include but are not limited to the items checked:

- ☐ BLACK BORDERS
- ☐ IMAGE CUT OFF AT TOP, BOTTOM OR SIDES
- ☐ FADED TEXT OR DRAWING
- ☐ BLURRED OR ILLEGIBLE TEXT OR DRAWING
- ☐ SKEWED/SLANTED IMAGES
- ☐ COLOR OR BLACK AND WHITE PHOTOGRAPHS
- ☐ GRAY SCALE DOCUMENTS
- ☒ LINES OR MARKS ON ORIGINAL DOCUMENT
- ☐ REFERENCE(S) OR EXHIBIT(S) SUBMITTED ARE POOR QUALITY
- ☐ OTHER: _____

IMAGES ARE BEST AVAILABLE COPY.

As rescanning these documents will not correct the image problems checked, please do not report these problems to the IFW Image Problem Mailbox.